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### Insect Pest Problem in Industrial Forest Plantation and Their Management

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#### ABSTRACT

*The problem in Industrial forest plantations confront from insect pests, with an emphasis on Indonesian *Acacia crassicaarpa* and *Eucalyptus* species. The reported insect pests *Glycyphana nicobarica*, *Helopeltis theivora*, *Altica* sp., and *Spodoptera exigua* are examined in this paper. Chemical procedures are not used as a last resort in management strategies, which prioritise environmental sustainability. Instead, they are integrated into cultural, mechanical, and biological controls, along with preventive measures. In order to stop and prevent insect pest infestations, this article promotes an integrated strategy that gives priority to environmental and economic concerns. This article supports sustainable practises in industrial forest plantations by highlighting balanced control strategies and proactive planning.*

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#### INTRODUCTION

The area of forest plantations will continue to grow and market demand will increase; by 2070, it is projected that forest plantations will expand to around 379 million ha (Nepal et al., 2019). One of the barriers or obstacles faced in forest plantations is pest attack. Pest attacks have

attacked *Acacia crassicaarpa* and *Eucalyptus* spp. since the seedling stage. Various reports have long reported this attack from various types of insect pests and from various insect orders, such as Lepidoptera, Hemiptera, Thysanoptera, and others (Kkadan et al., 2020).

In fact, many new insect pest attacks have been reported in forest plantations in several regions of Indonesia, including *Mylloceris scapularis* Roelofs (Coleoptera: Curculionidae) and *Rhytiphora bankii* (Fabricius) (Coleoptera: Cerambycidae). This pest attack occurs in *Acacia crassiparva* plantations in Riau and North Sumatra, Indonesia, with various levels of detrimental attacks. Attacks occur in nurseries up to 2 months old, and the visible symptoms of an attack are leaf damage caused by attacks by herbivore insects (Sirait et al., 2020).

There are many control options that can be used to overcome the problem of herbivorous insects, such as chemical and non-chemical applications. Although it cannot be denied that the forest plantation industry requires quick action to control insect pests, they tend to use chemical control. Although there are also many reports reporting the successful use of entomopathogens, bioinsecticides, natural enemies such as parasitoids and predators, resistant varieties, cultivation modification practices, and integrated pest management (IPM). The urgency of IPM can engineer the environment for the existence of various beneficial insects that are able to control the existence of herbivore insects. Dharma et al. (2018) found that organic cultivation efforts are able to increase the presence of parasitoids and predators in cultivated crops.

In an effort to control insect pests, it is necessary to know the economic threshold number of each type of pest insect at the point where pest attacks cannot be tolerated any longer, which can have fatal consequences if identification and control measures are too late. Control using chemicals is indeed the easiest to use, but there are many side effects produced, such as changes in the ecosystem, insect pest resistance, resuscitation, and killing beneficial insects. So preventive measures are also the most important part of efforts to manage the presence of insect pests in industrial forest plantations.

In this article, we will review the plants *A. crassiparva* and *Eucalyptus* spp., which are generally cultivated in industrial forest plantations, as well as the types of insect pests that are reported to attack these two plants, a discussion of pest control that has been empirically proven to be able to control them, and various control techniques that can be integrated with each other to prevent and suppress insect pest attacks by prioritising economic and environmental factors.

## FOREST PLANTATION CROPS

One of the trees used in industrial forest plantations is *Acacia crassiparva*, because it is known to have the potential to be developed in various ecosystem conditions and is usually cultivated in wet

land areas that are rich in organic material but minimal in minerals. *A. crassicarpa* is an endemic tree spread across Australia and Papua Island (Indonesia and Papua New Guinea), which has the ability to grow quickly and has strong wood and advantages for furniture, pulp and paper (Hanchor et al., 2016).

The strength of wood has an air-dry density of 710 kg per cubic metre and a basic density of 620 kg per cubic meter. *A. crassicarpa* is the best species of wood tree to be used as pulp and paper. Other advantages are the high nitrogen fixation ability and the rhizobium nodulation ability. The seeds of natural populations of *A. crassicarpa* originating from Papua New Guinea and Australia have been distributed in various countries in Asia, Africa, America, and various island countries such as the Leeward Islands, Hawaii, Micronesia, and Madagascar, although there are also many reports that not all areas are able to cultivate this tree (Pasieczni and McDonald, 2016).

*Eucalyptus* is a forest plant that is cultivated in industrial forest plantations apart from *Acacia crassicarpa*. The species commonly used is *Eucalyptus pellita*, which is used as raw material in the pulp and paper industry in the world. In Indonesia, for example, this species is used in mineral land areas. *E. pellita* has the ability to grow quickly and is usually harvested after 7

years; currently, it is even harvested at 5 years of age (Muhdi et al. 2019).

*E. pellita* is naturally distributed in Papua New Guinea, Australia, and Papua (Indonesia). Generally, *E. pellita* grows at an altitude of 0-800 m, with rainfall of 1000 to 3000 mm/year and a dry season of up to 5 months. For productivity, it is known that *E. pellita* will be more productive in areas with high rainfall. It is known that the productivity of *E. pellita* is between 9.4 and 34.9 m<sup>3</sup>/ha/year. not only in high rainfall areas, *E. pellita* is also able to grow in low rainfall (1000–1200 mm/year) and lower temperatures (22 °C), provided that many other factors will also influence its productivity, such as soil type, silvicultural practices, and season duration. Each of these factors will greatly influence; for example, a record increase of 100 mm of rainfall in Brazil can influence the biomass of the *E. grandis* × *E. urophylla* clone above ground to 2.3 Mg/ha/year (Hutapea et al., 2023)

## INSECT PESTS OF FOREST PLANTATION

At the beginning of the Industrial Plantation Forest (HTI) industry, the *Acacia mangium* species was the dominant species chosen for mineral and wet land types. However, this type is starting to be replaced by *Eucalyptus* sp. and *A. crassicarpa* due to several reasons, one of which is pest and disease attacks. However, currently, it cannot

be denied that insect pest attacks also frequently attack *E. pellita* and *A. crassicarpa* plants. Several reports of new insect pest attacks on industrial forest plantations have been reported.

Sirait et al. (2020) reported pest attacks on *A. crassicarpa* in North Sumatra, Indonesia, and also in Riau, Indonesia. It was reported that attacks by *Myllocerus scapularis* and *Rhytiphora bankii* were reported to attack many young plants in nursery and under-6-month-old plants. Previously, pests such as *M. scapularis* and

*R. bankii*, known by the common names Ash weevil and Longhorn beetle, were only reported on soybean and cotton plants in South Asian countries such as Sri Lanka and Bangladesh (Ali, 2008; Thakur et al., 2021). Next, in Table 1, several types of insect pests that often attack industrial forest plantations will be presented, as well as several new reports that have been recorded as attacking and causing harm to industrial forest plantations in the species *A. crassicarpa* and *E. pellita*.

**Table 1.** Major insect pest of industrial forest plantation

Forest Plantation Crop	Insect Pest	References		
	Common Name	Scientific Name		
	Common Name	Damage		
	Flower Beetle Cetoniinae	<i>Glycyphana nicobarica</i>	Damage caused on the leader shoot can create multiple leader shoots.	Sinulingga et al. 2020
	Tea Mosquito Bug	<i>Helopeltis theivora</i>		Roy et al. 2015
	Ash weevil	<i>Myllocerus scapularis</i>	Damage notching on leaf margins and defoliation leaving only the midrib.	Sirait et al. 2020
<i>Acacia crassicarpa</i>	Longhorn beetle	<i>Rhytiphora bankii</i>	Chews the branches of young.	
	Flea beetle	<i>Altica sp.</i>		Wilcox 1979,
	Red beetle	<i>Aulacophora sp.</i>		Tavares et al. 2022
	Leaf beetle	<i>Aulacophora coffeae</i>	Injuring and perforating the leaves.	
	Black-back cucumber beetle	<i>Aulacophora flavomarginata</i>		
	Beet armyworm	<i>Spodoptera exigua</i>		Sulistyono et al. 2020;
	Cutworm	<i>Spodoptera litura</i>	Damage of the leaves.	Khan et al. 2023.
	Termite	<i>Coptotermes curvignathus</i>	Destroy the wood or attacking trees, they	Haneda et al. 2017

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Termite	<i>Coptotermes gestroi</i>	eat the cellulose	
Termite	<i>Schedorhinotermes javanicus</i>	material.	
Termite	<i>Longipeditermes longipes</i>		
Termite	<i>Pericapritermes mohri</i>		
	<i>Euwallacea perbrevis</i>	They bore acacia stems and are associated with	Lynn et al. 2020
Ambrosia beetle	<i>Euwallacea fornicatus</i>	<i>Fusarium rekanum</i> sp. nov.	Tran et al. 2022; Hulcr et al. 2017
	<i>Eewallacea similis</i>		
Bagworm	<i>Pteroma plagiophleps</i>	Causes holes on the upper surface of the leaves, first yellow, then turning brown as if burned, and finally falling off.	Asmaliyah et al. 2019
Bagworm	<i>Pagodiella sp.</i>		Lelana et al. 2022
Mole cricket	<i>Gryllotalpa africana</i>	They feeds on young seedlings and low shoots.	Thu et al. 2010
Spiny moth	<i>Parasa pastoralis</i>	Larvae damage leaves. These insects eat wood, thus creating cavities in the wood and producing wood chips and fecal pellets or frass in quite large quantities.	Suka et al. 2022
Wood-feeding cockroach	<i>Panesthia angustipennis angustipennis</i>		Hawari et al. 2016
<i>Eucalyptus pellita</i>	Whiteflies	<i>Bemisia tabaci</i>	Chlorotic spotting, growth distortion, and premature leaf drop. Jufenlin et al. 2023
	Leafroller	<i>Strepsicrates macropetana</i>	Feeding on the shoot tips, buds and developing flowers, which are adhered together with webbing. Mauchline et al. 1999; Walker, 1866
	Horsfield's tussock moth	<i>Calliteara horsfieldii</i>	Damage of the leaves. Chung et al. 2015; Nair, 2001
	Geometer moth	<i>Hyposidra picaria</i>	Damage caused by the Leaf-cutting
	Nettle caterpillar	<i>Darna trima</i>	Caused necrotic, elongated holes surrounded by brown tissue.

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Green weevil	<i>Hypomeces squamosus</i>	Produce cuts, holes, or leaf falls.	
Leaf-cutting Bee	<i>Megachile</i> sp.	Damage caused by the Leaf-cutting.	Ruzzier et al. 2020; Akram et al. 2022
Termite	<i>Microcerotermes</i> sp.	Termites can attack and spread around trees, causes damage, and death of good at any stage of development from young or mature trees.	Prastyaningsih et al. 2020; Chung et al. 2015
Termite	<i>Discupiditermes nemorosus</i>		
Termite	<i>Termes rostratus</i>		
Termite	<i>Macrotermes gilvus</i>		
Termite	<i>Nasutitermes matangensis</i>		
Termite	<i>Schedorhinotermes medioobscurus</i>		
Termite	<i>Amitermes dentatus</i>		
Termite	<i>Coptotermes havilandi</i>		
Longhorned borers	<i>Phoracantha semipunctata</i>	Chewing through the frass plugs, Foliage can discolor and wilt, and limbs can die back.	Paine et al. 2009

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Longhorned borers	<i>P. recurva</i>		
The red coffee borer	<i>Zeuzera</i> spp.	They cause holes in the stem, swelling, breaking/forking, and/or bending.	Suheri et al. 2022
	<i>Polyphagozerra coffeae</i>		Tavares et al. 2020
Wood grasshopper	<i>Valanga nigricornis</i>	Usually eat leaves starting at the edges, while nymphs eat between the vein leaves creating holes in leaves.	Saputri et al. 2023
Asian ambrosia beetle	<i>Xylosandrus crassiusculus</i>	Beetles can cause tree death, because they associated with pathogenic fungi.	Thu et al. 2010
White grub	<i>Holotrichia trichophora</i>	Feeding on the shoot tips, buds and de on roots of young trees, girdling the stem below ground.	Thu et al. 2010
Scarab beetle	<i>Anomala</i> spp.	Causing a characteristic jagged, ripped pattern	Thu et al. 2010
Mole cricket	<i>Gryllotalpa africana</i>	They feeds on young seedlings and low shoots.	Thu et al. 2010
Guava moth	<i>Ophiusa disjungens</i>	Damage (defoliation) on Eucalyptus is up to 80.2% in Sumatra.	Raimon et al. 2020

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## MANAGEMENT OF INSECT PESTS OF INDUSTRIAL FOREST PLANTATION

Control of insect pests on forest plants is carried out both chemically and non-

chemically. In industrial forest plantations, treatment has even been carried out on the seeds that will be used as seedlings, and fungicide has been soaked on the seeds that

will be planted in the field. Control measures are not just actions after an insect pest attack occurs but also planning actions to prevent insect pest attacks, planned day by day and every month. because it is known that industrial forest plantations only cultivate homogeneous plants, so they are very susceptible to insect pest attacks.

#### ***Application of Synthetic Insecticides***

Synthetic insecticide applications are generally carried out in industrial plantation forests. Applications are carried out at plant age levels, starting with seedlings and 12-month-old plants. At the nursery stage, pest attacks on *A. crassicaarpa* and *E. pellita* were very diverse. Pesticide applications in industrial plantation forests use low-hazard insecticides because they are regulated by international regulations. Kkadan et al. (2020) conducted research on Acacia and Eucalyptus and found that the application of spinetoram was very effective in controlling *S. exigua* and thrips, with 100% and 95% effectiveness on each object tested using a spinetoram-based solution (0.12%).

Application of spinetoram to *Eucalyptus* also shows high effectiveness against attacks against *Strepsicrates semicanella*. Insecticide applications are also often carried out in various combinations to measure the level of effectiveness on insect pests, as was done by Saputra and Aluyah (2019). They tried to combine thuricide insecticide, which has the active ingredient *Bacillus thuringiensis*, with cassava tape suspension. In the application of insecticides to several insect pests, it was not only carried out using the spraying technique but also using the fogging technique. It was also found that, in some cases, the fogging technique was more effective in reducing pest attacks. Jufenlin et al. (2023) stated that the application of the insecticide Matador 250 EC with the active ingredient Lamda Sihalthrin using a spraying technique was able to reduce *B. tabaci* to 44% while the fogging technique was 56%, with the fogging application on *Strepsicrates macropetana* to 41% and to 26% on the spray technique.

**Table 2.** Some recommended synthetic insecticides for insect pest

<b>Crop</b>	<b>Synthetic Insecticide</b>	<b>Target Insect Pest</b>	<b>References</b>
A. <i>crassicaarpa</i>	Spinetoram	<i>S. exigua</i> ; <i>Scirtothrips sp</i>	Kkadan et al. 2020
	Chlorpyrifos	<i>S. exigua</i> ; <i>Scirtothrips sp</i>	
	Dimehipo	<i>X. Crassiusculus</i>	Saputra & Aluyah, 2019
	Amitraz		
	Piridaben	<i>Tetranychus Sp.</i>	Lumbantobing et al. 2023
	Propargite		
	Sulfur		



<i>E. Pellita</i>	Spinoteram	<i>Strepsicrates semicanella</i>	Kkadan et al. 2020
	Lamda Sihalotrin	<i>Bemisia tabaci; Strepsicrates macropetana</i>	Jufenlin et al. 2023
	Siantraniliprol + Lufenuron		
	Imidakloprid	<i>Strepsicrates sp</i>	Agustin et al. 2023
	Permetrin + Abamektin		
	Dinotefuran		
	Tefluthrin		
	Fonotos		
	Fipronil	Various Termite	Wilcken et al. 2002
	Disufolton + Triadimenol		
Acephate			

**Cultural Control**

Adopting appropriate silvicultural practices to prevent insect pests has been implemented in industrial forest plantations. Appropriate silviculture practices have been chosen as preventive measures against insect pest attacks, so that they become a holistic unit in preventive action steps. The practical practices that can be considered are as follows:

**Land Preparation**

In forest industrial plantings, one of the typical technical practises is land preparation. Tilling the soil generally aims to loosen the soil mass to allow sufficient room for the growth and development of plant roots in the soil. One of the things that can affect the emergence of disease and insect attacks is the preparation of the land. This procedure has the ability to control weeds in addition to influencing the presence of diseases and pests. According to Nuri (2005), the best way to prevent white root rot disease

is to prepare the site mechanically. This is because machinery can remove stumps and leftover roots from the soil.

**Plant Population Manipulation**

Planting distances for *A. crassicarpa* and *E. pellita* are regulated in such a way, not only guided by quantitative wood production, that research results show that planting distances have a significant effect on growth in height and diameter, and planting distances do not have a significant effect on survival percentage and competition index. 12 month-old plants (Purwanti et al., 2022). Plant spacing is also considered a preventive measure to control insect pests. *A. crassicarpa*, which is cultivated in peat areas, has a planting distance of 3 x 2 metres, while *E. pellita*, which is generally cultivated in mineral soil, has a planting distance of 3 x 2 and also has other options such as 3 x 2.5 m, 3.5 x 2 m, and 3.3 x 2.2 m. Asmaliyah & Rostiwati (2015) stated that a planting distance of 3 x 4

m is an effective planting distance for suppressing the development of the *Cycnotrachelus* sp. beetle pest and the pathogen *Cephaleuros* sp. on land island plants in the third year after planting. This condition is indicated by a decrease in the percentage of attacks by 0.57% and a reduction in the intensity of attacks by beetle pests by 2.48% and pathogenic diseases by 8.44%.

#### *Supplemental Application of Soil Nutrients*

In addition to determining a plant's resistance to several pest and disease attacks, good soil has a significant role in plant yield (Herliyana et al. 2012). In industrial plantation forests, organic materials such as ameliorant materials obtained from factory processing residues are used in conjunction with additional nutritional supplements and fertilisers to increase soil fertility. These materials improve the physical, chemical, and biological conditions of the soil. In industrial planted forests, almeiron material is typically used prior to *A. crassicarpa* planting on peatlands. You must be careful when applying the right type and amount of fertiliser, though, as some fertilisers have the potential to exacerbate pest assaults.

#### *Weed Control*

Pest attacks in industrial plantation forests are not the only problem; another problem that will be faced is the presence of weeds, because they will become competitors for the main plants for sunlight,

nutrients, water, and space to live (Wibowo, 2006). It is well known that several types of weeds that are similar to the main crop usually become nests for several insect pests (Capinera, 2005). In plantations around *A. crassicarpa*, there are generally many other types of acacia that are not cultivated, such as *A. mangium*, which has been studied and can be a place for insect pests to colonize. So by understanding proper weed control, we can suppress the presence of insect pests, and unwanted weeds such as *A. mangium* can also become a diversionary plant from the main crop.

#### *Trap Cropping*

One way to combat insect pests is through cultural control methods such as trap plants, which are plants that are planted alongside or ahead of the primary plant. By concentrating pests in one area, trap crops can improve the effectiveness of pest control. Biological control agents can also be released into trap crops, where they serve as a haven for beneficial organisms that eventually spread to the main crop (Reddy, 2017). When crop diversification and the reduction of synthetic inputs are given priority over crop output alone, trap crops can do particularly well on lower production settings.

#### *Use of Bioinsecticide*

Natural substances like pheromones, plant bioactive chemicals, and microorganisms like bacteria, fungus,

viruses, or protozoa are the sources of bioinsecticides. Numerous chemical defence mechanisms against insects are known to exist in plants. Plants possess phytochemicals that serve as insect repellents, toxins, feeding deterrents, and growth regulators.

The entire body of small plants, a combination of different plants, or sections of higher plants can be utilised as components to manufacture bioinsecticides or botanical insecticides. (Demirak Şengül & Canporat, 2022). Rusandi et al. (2016) stated that mahogany seeds have the potential to control *S. litura* on *A. crassiparva*; giving mahogany seed extract at a concentration of 30 g/l of water is the best concentration for controlling *Spodoptera litura* F., with the fastest initial death time of 2.75 hours. (2 hours, 45 minutes), the fastest time of death is 2 hours, 45 minutes, and there are many other plants that can be used as biopesticides.

#### *Use of Insect Resistant Varieties*

Planting plant clones that are fully or partially resistant to pest attacks is an effective way to reduce damage to industrial forest plantations. so that currently many forestry companies are developing clones that are resistant to pests and diseases (Hutasuhut et al. 2015). Resistant clones are obtained from the selection of certain individuals with characteristics in accordance with the purpose of breeding, so that a breeding population is formed. From the

breeding population, further selection can be carried out to obtain individuals with certain advantages, which can then be crossed or hybridised to produce new individuals with combined characteristics from both parents (Sunarti, 2018).

#### ***Mechanical Control***

Mechanical control is physical pest control, namely the direct treatment of plants using certain tools or manually. This technique relies on real action, in the form of using tools to ward off animals and diseases. Mechanical techniques do not rely on chemicals such as insecticides or drugs. Instead, they use tools such as handpicks, sickles, sticky yellow traps, lamp traps, plant scissors, machetes, and so on. According to Dudley et al. (2012), one mechanical engineering procedure involves treating the seeds, which involves cleaning them of any foreign things and sealing them in plastic bags that are laminated and contain carbon dioxide for a period of two weeks before storing them.

To prevent contamination, methyl bromide is also applied to the seeds. Additional protection against insect pest attacks comes in the form of hot water and acid scarification techniques to break dormancy. However, when insect larvae are isolated in big seed treatments, these procedures might not be entirely effective. Cans, jute bags, and baskets can all be used to store seeds.

The use of pheromone traps to regulate *Zeuzera* sp. imago in *E. pellita* is one type of mechanical control that has been implemented in industry. *Zeuzera* sp. is responsible for broken lower portions of the plant, swollen stems, and broken upper crowns, according to Suheri et al. (2020). The study he carried out produced encouraging findings. The installation of delta traps and *Zeuzera* pheromones pyrene was used to manage pests. Still, the outcomes are insufficient, necessitating comprehensive pest management.

### **Biological Control**

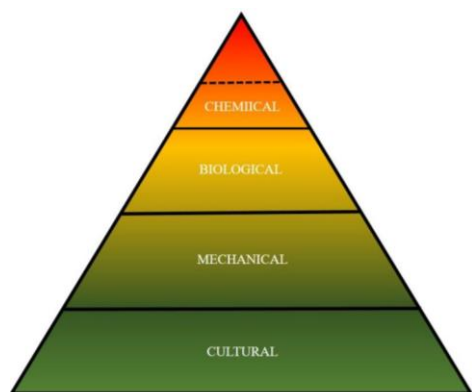
Biological control is currently getting more and more popular as people realise how crucial it is to preserve sustainability and environmental balance. The truth is that, independent of the environment, natural creatures regulate approximately 98% of pest populations. Due to their persistent nature, ongoing buildup in the environment, toxicity to biotic components of life, and potential health risks to humans, chemical pesticides and insecticides are used far less frequently these days.

Utilising beneficial insects, such as parasitoids and predators, that are either naturally occurring in the environment or purposefully released to manage insect pests, is known as biological control. Pathogens, such as fungi and viruses, can also be used to manage undesirable insects. The entomopathogenic fungus *Beauveria bassiana*

and *Lecanicillium lecanii*, which both have significant mortality capacity but are in different life stages of insect pests, can control *Helopeltis* sp., one of the major pests on *Acacia*. Laboratory research revealed that the entomopathogenic fungus *B. bassiana* was successful in causing 100% mortality of *Helopeltis* sp. imago at conidia density of  $10^8$  and  $10^9$ /mL, while the entomopathogenic fungus *L. lecanii* caused 100% mortality of third instar nymphs of *Helopeltis* sp. at a conidia density of  $10^9$ /mL (Anggarawati et al. 2017).

*Sycanus* sp. is a naturally occurring predatory insect that is commonly found in peatland areas. It is the natural enemy of several insect pests of *A. crassicaarpa*. Interestingly, the presence of Geronggang (*Cratoxylon arborescens*), a native tree species in peat areas that serves as a habitat for *Sycanus* sp., can maximise the potential of *Sycanus* sp. (Suchayono et al. 2022). *S. litura*, which is a dangerous polyphagous pest in *A. crassicaarpa* and *E. pellita* nurseries, has been able to be controlled by utilising the nuclear polyhedrosis virus (NPV) (Gafur et al. 2012) (Yasin et al. 2020).

### ***Integrated Pests Management in Industrial Forest Plantation***



**Figure 1.** Pyramid of Pest Management Tactics

Integrated Pest Management (IPM) is a strategy used by nature to control pests by integrating various types of control (**Figure 1**), such as cultural, mechanical, and biological. which aims to be environmentally friendly and sustainable. IPM has been widely carried out in industrial forest plantations on *A. crassiparpa* and *E. pellita* plantations. In research conducted by Saputra and Aluyah (2019), they combined cassava extract with insecticides to control Ambrosia pests on *A. crassiparpa* plants and got effective results. Sucahyono et al. (2022) conducted research on the effect of *C. arborescens* plants on the release of *Sycanus* sp., which is a natural enemy for many insect pests in industrial forest plantations, especially peatland areas.

IPM strives for both temporal and spatial sustainability, according to Luo and Shen (1998). The goal is to focus on natural preventative and control methods and to always act cautiously when implementing

any control techniques that may have negative environmental effects. When the pest has surpassed the economic threshold, this has to be given priority as well. As a matter of fact, IPM has emerged as a benchmark for the management of industrial plantation forests, bolstered by numerous rules that prioritise the preservation of the ecosystem. The non-governmental organisation Forest Steward Council (FSC) forbids the use of specific chemicals in insect pest management programmes (Lemes et al., 2017).

### **CONCLUSION**

There are many insect pests that cause damage and losses to industrial plantation forests, starting from seedlings and also after being transplanted into the field. These pests attack at various stages of insect life. However, there are many solutions that can be used to control these pests, and IPM seeks to integrate each type of control, excluding the application of chemical insecticides, which are highly effective but are feared to produce dangerous environmental impacts such as pest resistance and also changes in the behaviour of other insect pests. There are many things that need to continue to be developed, such as combining various types of control, such as continuing to develop pest-resistant clones, using entomopathogens, and so on.

## REFERENCES

- Agustin, Syahrani, Agus Priyono, dan Karti Rahayu Kusumaningsih. (2023). Uji Efektivitas Beberapa Jenis Insektisida Terhadap Pengendalian Hama Ulat Penggulung Daun (*Strepsicrates* sp.) Pada Bibit Eucalyptus Hybrid. Jurnal Mahasiswa Instiper (AGROFORETECH) 1 (1):810-15.
- Anggarawati, S. H., Santoso, T., Anwar, R. (2017). The Use of Entomopathogenic Fungi *Beauveria bassiana* (Balsamo) Vuillemin and *Lecanicillium lecanii* (Zimm) Zare & Gams) for Controlling *Helopeltis antonii* Sign (Hemiptera: Miridae). Jurnal Silviculture Tropika Vol. 08 No. 3, Desember, Hal 197-202.
- Ali, M. I. (2008). A survey of the insect pests of soybean in Northern Bangladesh, their damage and occurrence. Tropical Pest Management Volume 34, 1988 - Issue 3. <https://doi.org/10.1080/09670878809371266>.
- Akram, W., Sajjad, A., Ghramh, H. A., Ali, M., Khan, K. A. (2022). Nesting Biology and Ecology of a Resin Bee, *Megachile cephalotes* (Megachilidae: Hymenoptera). Insects. 13(11):1058. <https://doi.org/10.3390/insects13111058>
- Asmaliyah, Hadi, E. E., Irianto, R. S. B. (2019). Pests and diseases on forest plant in burned peatlands in South Sumatra. IOP Conf. Ser.: Earth Environ. Sci. 308 012071.
- Asmaliyah, A., Rostiwati, T. (2015). Pengaruh Pengaturan Jarak Tanam Terhadap Perkembangan Serangan Hama Dan Penyakit Pulai Darat (*Alstonia angustiloba*). Jurnal Penelitian Hutan Tanaman Vol. 11 No. 3, April : 41-50.
- Capinera, J.L. (2015). Relationships between insects and weeds: An evolutionary perspective. Weed Sci. 53, 892–901.
- Chung, A., Kimjus, K., Ajik, M., Ong, R., Johnlee, E. (2015). A Note On Some Pests Of *Eucalyptus* In Sabah, Malaysia. 10.13140/RG.2.2.17339.26401.
- Dharma, T. A., Sitepu, S. F., Lubis, L., & Br. Girsang, S. S. (2018). Kelimpahan Serangga Penghuni Tajuk pada Pertanaman Bawang Merah Semi Organik dan Konvensional. Jurnal Pertanian Tropik , 5(2), 268- 275. <https://doi.org/10.32734/jpt.v5i2.3012>.
- Dudley, T. L., Bean, D. W. (2012). Tamarisk biocontrol, endangered species risk and resolution of conflict through riparian restoration. Biocontrol 57:331–347.
- Gafur, A., Nasution, A., Tarigan, M., Tjahjono, B. (2012). Development Of Biological Control Agents To Protect Plantation Forests In Sumatra, Indonesia. Proceeding International Conference on the Impact of Climate Change to Forest Pests and Diseases in The Tropics. UGM, Indonesia. Hal 200-206.
- Hanchor, U., S. Maelim, W. Suanpaga, J. M. Park, and K. S. Kang. (2016). Growth performance and heritability estimation of *Acacia crassicarpa* in a progeny trial in eastern Thailand. Silvae Genetica 65(2): 58–64. DOI: 10.1515/sg-2016-0017.
- Haneda, N. F., Retmadhona, I. Y., Nandika, D., Arinana. (2017). Biodiversity of subterranean termites on the *Acacia crassicarpa* plantation. Biodiversitas. Volume 18, Number 4. Pages: 1657-1662.
- Hawari, S. Y., Muhammad, A., Salbiah, D. (2016). Preferensi Lipas Kayu (*Panesthia angustipennis*) Terhadap Kayu Akasia (*Acacia Crassicarpa*) Dengan Tingkat Lapuk Berbeda. Repository Unri. <http://repository.unri.ac.id/xmlui/handle/123456789/7782>.

- Herliyana, E. N., Achmad, Putra, A. (2012). Liquid Organic Fertilizer Influence on Jabon (*Anthocephalus cadamba* miq.) Seedling Growth and Its Resistance to Disease. *Jurnal Silviculture Tropika*. Vol. 03 No. 03, Hal. 168 – 173.
- Hulcr, J., Black, A., Prior, K., Chen, C. Y., Li, H. F. (2017). Studies of Ambrosia Beetles (Coleoptera: Curculionidae) in Their Native Ranges Help Predict Invasion Impact. *Florida Entomologist*, 100(2), 257-261. <https://doi.org/10.1653/024.100.0219>
- Hutapea, F. J., Christopher J. W., Mendham, D., Volkova, L. (2023). Sustainable management of *Eucalyptus pellita* plantations: A review, *Forest Ecology and Management*, Volume 537. <https://doi.org/10.1016/j.foreco.2023.120941>.
- Hutasuhut, J. H., Anna, M., Siregar, E. B. M. (2015). Uji Infeksi *Cylindrocladium* SP. Pada Klon Hibrid *Eucalyptus Grandis* X *Eucalyptus Urophylla* (Infection Test *Cylindrocladium* SP. on Hybrid Clones of *Eucalyptus Grandis* X *Eucalyptus Urophylla*). *Peronema Forestry Science Journal*, vol. 4, no. 3, pp. 143-148.
- Jufenlin, M., Priyono, A., Sumardi. (2023). Uji Efektivitas Metode Semprot dan Pengkabutan terhadap Intensitas Serangan Hama Whiteflies (*Bemisia tabaci*) dan Leafroller (*Strepsicrates macropetana*) pada Tanaman Induk *Eucalyptus* Spp. di Indoor Breeding Orchard (Ibo) Kerinci Research Nursery. *Agroforetech*. Volume 1, Nomor 01.
- Khan, S., Duran, a., Ikram, M., Sinulingga, N. G. H., Tavares, W. D. S., Sirait, B. A., Kkadan, S. K., Tarigan M. (2023). *Trichogramma yousufi* sp. nov. Employed for the Management of *Spodoptera exigua* and *Spodoptera litura* in Indonesia. *Florida Entomologist*, 103 (3) : 353-359. <https://doi.org/10.1653/024.103.0307>.
- Kkadan, S. K., Sirait, B. A., Asfa, R., Tavares, W. D. S. Tarighan, M., Duran, A., Wong, C.Y, Sharma, M., (2020). Evaluation of a spinetoram-based insecticide against *Y. lepidopteran* and thrips infesting *acacia* and *eucalyptus* in Sumatra, Indonesia. *Journal of Entomology and Zoology Studies* 8(2): 1345-1351.
- Lelana, N. E., Utami, S., Darmawan, U. W., Nuroniah, H. S., Darwo, Asmaliyah, Haneda, N. F, Arinana, Darwiati, W., Anggraeni, I. (2022). Bagworms in Indonesian Plantation Forests: Species Composition, Pest Status, and Factors That Contribute to Outbreaks. *Diversity*. 14, 471. <https://doi.org/10.3390/d14060471>.
- Lemes, P. G., Zanuncio, J. C., Serrão, J.E. *et al.* (2017). Forest Stewardship Council (FSC) pesticide policy and integrated pest management in certified tropical plantations. *Environ Sci Pollut Res* **24**, 1283–1295. <https://doi.org/10.1007/s11356-016-7729-3>.
- Lumbantobing, I. R. C., Wijayani, S., & Andayani, S. T. (2023). Uji Efektivitas Beberapa Akarisida untuk Pengendalian Hama Tungau Merah (*Tetranychus* Sp.) pada Tanaman Induk *Acacia Crassicarpa*. *Agrotechnology, Agribusiness, Forestry, and Technology: Jurnal Mahasiswa Instiper (AGROFORETECH)*, 1(1), 796–800.
- Lynn, K. M. T., Wingfield, M. J., Duran, A., Marincowitz, S., Oliveira, L. S. S., de Beer, Z. W, Barnes, I. (2020). *Euwallacea perbrevis* (Coleoptera: Curculionidae: Scolytinae), a confirmed pest on *Acacia crassicarpa* in Riau, Indonesia, and a new fungal symbiont; *Fusarium rekanum* sp. nov. *Antonie van Leeuwenhoek*. <https://doi.org/10.1007/s10482-020-01392-8>
- Mauchline, N. A., Withers, T. M., Wang, Q., Davis, L. (1999). Life history and abundance of the *Eucalyptus* leafroller

- (*Strepsicrates macropetana*). Proc. 52nd N.Z. Plant Protection Conf. 108-112.
- Muhdi, Sahar, A., Hanafiah, D. S., Zaitunah, A., Nababan, F. W. B. (2019). Analysis of biomass and carbon potential on eucalyptus stand in industrial plantation forest, North Sumatra, Indonesia. IOP Conf. Ser.: Earth Environ. Sci. 374 012054.
- Nair, K. S. S. (2001). Pest Outbreaks In Tropical Forest Plantations: Is There a Greater Risk for Exotic Tree Species?. Center for International Forestry Research
- Nepal, P., Korhonen, J., Prestemon, J. P., Cabbage, F. W. (2019). Projecting global planted forest area developments and the associated impacts on global forest product markets. J. Environ. Manag. 240, 421–430. <https://doi.org/10.1016/j.jenvman.2019.03.126>.
- Nuri, S. F. (2005). Dampak Penyiapan Lahan Acacia crassicarpa Terhadap Serangan Penyakit Busuk Akar Putih. Theses. IPB University. Bogor
- Paine, T. D., Dreistadt, S. H., Millar, J. G. (2009). Pest Notes: *Eucalyptus longhorned* Borers. UC Statewide IPM Program, University of California, Davis, CA 95616.
- Pasieczni, N., McDonald, D. (2016). *Acacia crassicarpa* (northern wattle). CABI Compendium. <https://doi.org/10.1079/cabicompendium.2192>
- Prastyaningsih, S. R., Hardiwinoto, S., Musyafa, Koranto, C. A. D. (2020). Diversity of termites (Isoptera) on industrial forest plantation of *Eucalyptus pellita* stands of tropical ecosystem in Riau, Indonesia. Biodiversitas. 21 (11): 5498-5505.
- Purwanti, E., Nurjanto, H. H., Widiyanto. (2022). Pertumbuhan Acacia Crassicarpa Umur 12 Bulan Pada Berbagai Jarak Tanam Di KHDTK Wanagama. Undergraduate Theses. Universitas Gadjah Mada. Yogyakarta.
- Ramon, A. L., Sinulingga, N. G. H. B., Ferlianda, I. A., Samosir, M. N., Pane, S. B., Kkadan, S.K., Tavares, W. de S., Tarigan, M., Duran, A. (2020). First report of *Ophiusa disjungens* (Walker, 1858) on *Acacia mangium* Willd. (Fabaceae), and damage and notes of its biology on *Eucalyptus* (Myrtaceae) commercial plantations in Sumatra, Indonesia (Lepidoptera: Erebidae). SHILAP Revta. lepid., 48 (191) : 439-447.
- Reddy, P. (2017). Trap Cropping. Agro-ecological Approaches to Pest Management for Sustainable Agriculture.
- Roy, S., N. Muraleedharan, A. Mukhapadhyay, and G. Handique. (2015). The tea mosquito bug, *Helopeltis theivora* Waterhouse (Heteroptera: Miridae): its status, biology, ecology and management in tea plantations. International Journal of Pest Management 61: 179–197.
- Rusandi, R., Mardhiansyah, M., Arlita, T. (2016). Pemanfaatan Ekstrak Biji Mahoni sebagai Pestisida Nabati untuk Mengendalikan Hama Ulat Grayak (*Spodoptera litura* F) pada Pembibitan Acacia Crassicarpa A. Cunn. Ex Benth. Jurnal Online Mahasiswa Fakultas Pertanian Universitas Riau, vol. 3, no. 1, Feb. pp. 1-7.
- Ruzzier, E., Menchetti, M., Bortolotti, L., Selis, M., Monterastelli, E., Forbicioni, L. (2020). Updated distribution of the invasive *Megachile sculpturalis* (Hymenoptera: Megachilidae) in Italy and its first record on a Mediterranean island. Biodiversity Data Journal 8: e57783. <https://doi.org/10.3897/BDJ.8.e57783>.
- Saputra, C. Dan Aluyah C. (2019). Pengaruh Dosis Suspensi Tape Singkong Dan Jenis Insektisida Dalam Mengendalikan Kumbang Penggerek Batang Ambrosia Pada Tanaman



- Krasikarpa (*Acacia Crassicarpa*) Di Pt. Bumi Mekar Hijau. Sylva. VIII – 1.
- Saputri, Y. A., Indriyanto, Asmarahman, C. (2023). Densitas Hama Bibit Tanaman Hutan Di Persemaian Permanen Bpdashl Way Seputih Way Sekampung Lampung Selatan. Jurnal Belantara Vol. 6, No.2, Agustus (191-203).
- Şengül Demirak MŞ, Canpolat E. (2022). Plant-Based Bioinsecticides for Mosquito Control: Impact on Insecticide Resistance and Disease Transmission. Insects. Feb 3;13(2):162.
- Luo Y. Q., Shen R, X. (1998). Sustainable pest management in forest (SPMF). Journal of Beijing Forestry University 20:96–98.
- Sinulingga, N. G. H., Grace, N. Suka, D. C. H., Sibuea, P., Tavares, W. D. S., Kkadan, S. K., Tarigan, M. (2020). *Glycyphana nicobarica* Janson (Coleoptera: Scarabaeidae: Cetoniinae): First Report on *Acacia crassicarpa* A. Cunn. ex Benth (Fabaceae) and Its Attraction to Water Softeners in Riau, Indonesia. The Coleopterists Bulletin, 74(1) : 62-64. <https://doi.org/10.1649/0010-065X-74.1.62>.
- Sirait, B. A., Sinulingga, N. G. H., Samosir, M. N., Tavares, W. D. S., Kkadan, S. K., Tarigan M., Duran, A. (2020). First Report Of *Mylocerus scapularis* Roelofs (Coleoptera: Curculionidae) And *Rhytiphora bankii* (Fabricius) (Coleoptera: Cerambycidae) On Commercial Plantings Of *Acacia crassicarpa* (Fabaceae) In Indonesia. The Coleopterists Bulletin, 74(2): 404–407.
- Sucahyono, M. P., Hidayat, R., Gafur, A., Indrayadi, H., Wahno, I., Herdyantara, A. B., Tjahjono, B. (2022). The potential of geronggang (*Cratoxylon arborescens* (Vahl.) Blume) in conserving *Sycanus* sp. as natural enemy of insect pests in peatland areas. IOP Conf. Ser.: Earth Environ. Sci. 1025 012041
- Suheri, M., Haneda, N. F., Anwar, R., Jung, Y., Sukeno, S., Park, J. (2022). Population dynamics of *Zeuzera* spp. (Lepidoptera: Cossidae) on *Eucalyptus pellita* plantation in Central Kalimantan, Indonesia. Biodiversitas. Volume 23, Number 11. Pages: 5782-5789.
- Suheri, M., Haneda, N. F., Jung, Y. H., Sukeno, S., Moon, H. K. (2020). Effectiveness of pheromone traps for monitoring *Zeuzera* sp. (Lepidoptera: Cossidae) population on *Eucalyptus pellita* plantation. IOP Conf. Ser.: Earth Environ. Sci. 468 012016
- Suka, D. C. S., Br Sinulingga, N. G. H., Wardoyo, A. S., Sirait, B. A., Sihombing, I. F. L., Tavares, W. D. S., Kkadan, S. K., Tarigan, M., Duran, A., Solovyev, A. V. (2022). First Report of *Parasa pastoralis* (Lepidoptera, Limacodidae) on *Acacia crassicarpa* (Fabaceae), and *Sycanus collaris* (Hemiptera, Reduviidae) as Its Predator in Sumatra, Indonesia. The Journal of the Lepidopterists' Society 76(1), 83-86. <https://doi.org/10.18473/lepi.76i1.a9>.
- Sulistiyono E, Kkadan SK, Maretha MV, Tavares W de S, Sirait BA, Sinulingga NGHB, Tarigan M, Duran A. (2020). First report, morphological and molecular identification of Spodoptera species (Lepidoptera, Noctuidae) on *Acacia crassicarpa* (Fabaceae) in Sumatra, Indonesia. Journal of the Lepidopterists' Society 74. <https://doi.org/10.18473/lepi.74i3.a4>
- Sunarti, S. (2018). The role of biodiversity in forest plant breeding: A case study on the development of new *Acacia* hybrid varieties (*Acacia mangium* x *Acacia auriculiformis*). Pros Sem Nas Masy Biodiv Indon Volume 4, Nomor 1, Hal 28-34.
- Tavares, W. D. S., Suka, D. C. G., Hardi, D., Adriya, R., Sinulingga, N., G. H., Kkadan, S. K., Tarigan, M., Duran, A.

- (2020). *Acacia crassicarpa* A. Cunn. ex Benth. (Fabaceae) as a Host Plant of Five Species in the Genera *Altica* Geoffroy, *Aulacophora* Dejean, and *Aulonogria borchmanni* (Coleoptera: Chrysomelidae, Tenebrionidae) in Sumatra, Indonesia. The Coleopterists Bulletin, 74(4) : 782-785. <https://doi.org/10.1649/0010-065X-74.4.782>.
- Tavares, W. D. S., Kkadan, S. K., Hendrik, A. M., Tarigan, M., Asfa, R., Yakovlev, R. V., Tachi, T., Duran, A., Wong, C. Y., Sharma, M. (2020). Notes on the biology and natural enemies of *Polyphagozerra coffeae* (Nietner, 1861) infesting *Eucalyptus pellita* F. Muell. (Myrtaceae) trees in Riau, Indonesia (Lepidoptera: Cossidae, Zeuzerinae). SHILAP Revta. lepid., 48 (190) junio ; 333-349.
- Thakur, K., Sharma, A., Sharma, K. (2021). Management of agricultural insect pests with physical control methods. The Pharma Innovation Journal; SP-10(6): 306-314.
- Thu, P. Q., Griffiths, M. W., Pegg, G. S., McDonald, J. M., Wylie, F. R., King, J., Lawon, S. (2010). Healthy plantations: a field guide to pests and pathogens of Africa, Eucalyptus and Pinus in Vietnam. Department of Employment. Economic Development and Innovation. Queensland. Australia.
- Tran, H., Thu, Q. P., Chi, N., M., Dell, B. (2022). Impacts and trapping of ambrosia beetles *Euwallacea fornicatus* and *E. similis* in Acacia plantations in Vietnam. Southern Forests: a Journal of Forest Science. 84.
- Wibowo, A. (2006). Gulma di Hutan Tanaman dan Upaya Pengendaliannya. Pusat Penelitian dan Pengembangan Hutan Tanaman. Badan Penelitian dan Pengembangan Kehutanan. Bogor.
- Wilkin, C. F., Raetano, C. G., Forti, L. C. (2002). Termite Pests in Eucalyptus Forest of Brazil. Sociobiology. Vol: 40 No. 01.
- Wilcox, J. A. (1979). Leaf Beetle Host Plants in Northeastern North America (Coleoptera: Chrysomelidae). North American Beetle Fauna Project, World Natural History Publications, Kinderhook, NY, 30 pp.
- Yasin, M., Qazi, M.S., Wakil, W. et al. (2020). Evaluation of Nuclear Polyhedrosis Virus (NPV) and Emamectin Benzoate against *Spodoptera litura* (F.) (Lepidoptera: Noctuidae). Egypt J Biol Pest Control 30, 88. <https://doi.org/10.1186/s41938-020-00271-8>.